

Setup for Live Imaging using the HCF1 Microscope

Please ask for help if you are unsure

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General considerations

Choice of Media

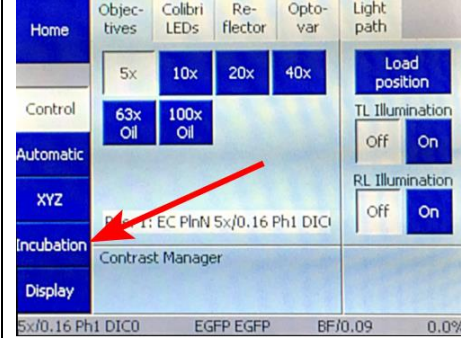
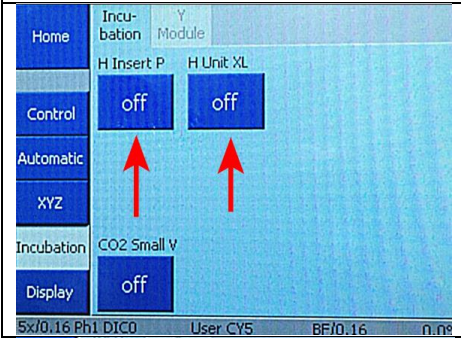
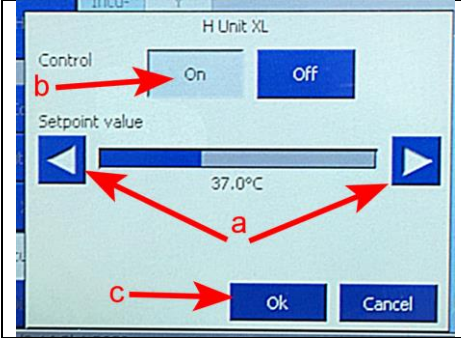
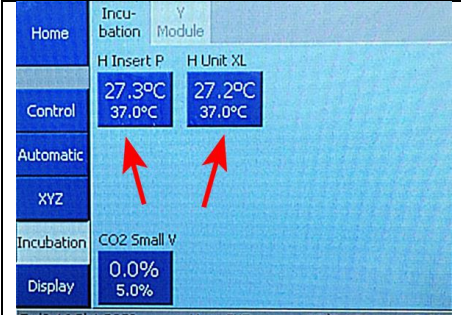
- Phenol Red dye in culture media has low-level auto-fluorescence particularly in the green emission wavelengths, which can interfere with the fluorescent signal, and is best avoided. There are Phenol Red free media available for fluorescent imaging.

Well Liquid levels

- Imaging for several hours or more at 37°C can cause evaporation of the culture/imaging media. Make sure each well has sufficient liquid level to compensate for this. Also filling unused wells in the same plate can be beneficial. Placing a container of water at the base of the microscope incubation chamber helps to provide a humid environment (an old pipette box with about 1cm of water is usually sufficient).

Heating

The HCF1 can provide heating to both the chamber (H Unit XL) and also to a heated stage insert for 35mm dishes (H Insert P – see Stage Inserts). Each unit can be set to a specified temperature on the TFT control screen. Switch on at least 30 minutes before use to allow the chamber to reach temperature and equilibrate. For longer term imaging (time series) then pre-heat the chamber for at least 2 hours before use, to prevent focus drift).

	<p>Turning on</p> <ul style="list-style-type: none"> Start up the microscope in the usual way Make sure all incubator doors are closed. On the TFT screen, select “Microscope” and then “Incubation” (A) (if Incubation is not visible then re-start the system).
	<ul style="list-style-type: none"> The display will show the temperature of the H unit XL and the H Insert P as OFF To Switch on, press the OFF button to open the temperature controls for the required unit(s).
	<p>Adjust the temperature:</p> <ul style="list-style-type: none"> Select the required temperature (a) and then press ON (b). Then press the OK button (c).
	<ul style="list-style-type: none"> The display will now show the current and the required set temperatures of each unit. Press the Control button to return to the objective display

Humidity

Imaging of live samples will result in evaporation of the growth media supporting the cells. This will become a significant problem for longer term imaging at higher temperatures (eg 37°C). This evaporation can be significantly reduced by placing a container (a base of a used pipette tip box is ideal) partially filled (1cm deep) with water inside the base of the microscope incubation chamber but away from the stage movement. If running over several days this box should be checked and filled daily.

Take care not to spill water inside the microscope incubator chamber

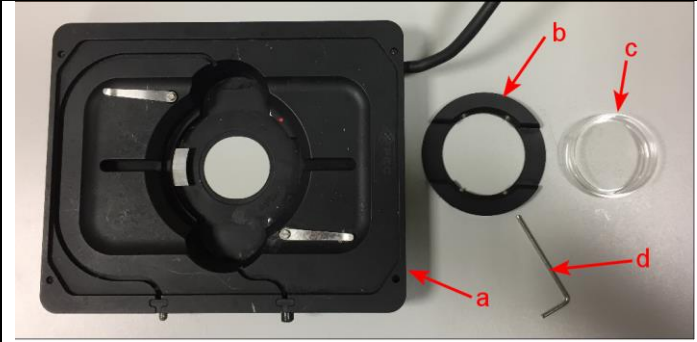

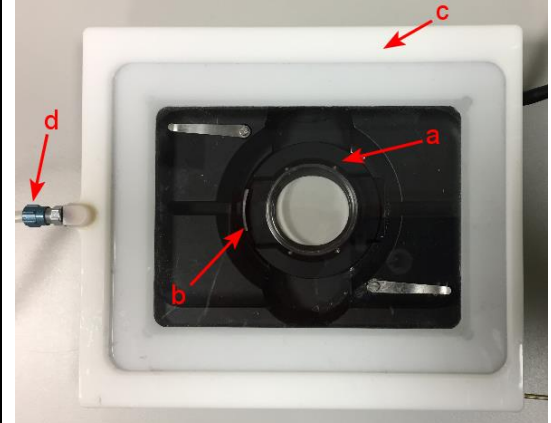
CO₂

Please note that the microscope incubator chamber, as a whole unit, cannot maintain a controlled CO₂ environment. Instead, it is possible to supply a CO₂ to a specific sample carrier lid. There are several lid/insert options for 35mm dishes, chamber slides and multi-well plates.

Stage Inserts

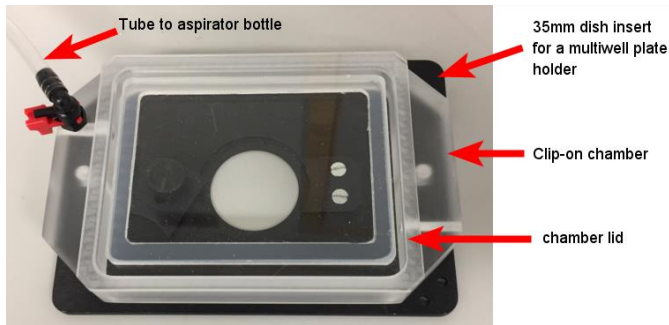
H Insert P

This is a heated stage insert that replaces the standard slide of multi-well plate inserts

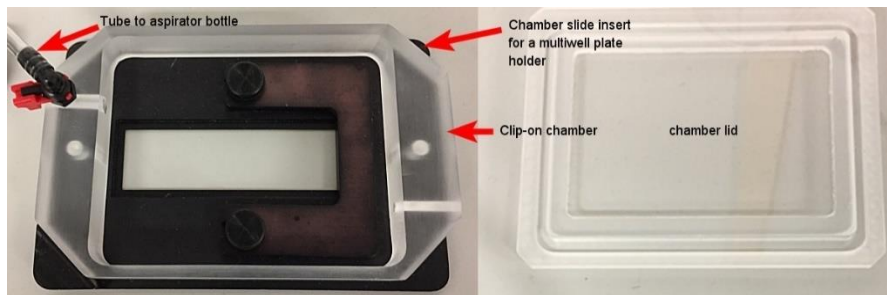
	<ul style="list-style-type: none">a. Heated stageb. 35mm dish ring clampc. 35mm dish Alan keyd. Spring plate
	<p>Loosen the screws on the ring clamp and on a flat surface, place the 35mm dish into the clamp. Tighten screws to just hold the dish (DO NOT OVER TIGHTEN).</p>
	<p>Inset the clamp and dish (a) into the stage by sliding the clamp against the spring plate (b). Place the glass lid on top. Attach the tubing to the CO₂ aspirator bottle.</p>

Other options

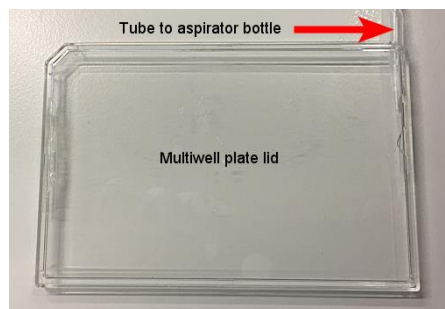
- 35mm dish and slide assembly unit that fits the multi-well plate holder



- Chamber slide



- Multi-well plate lid for normal multiwell plates

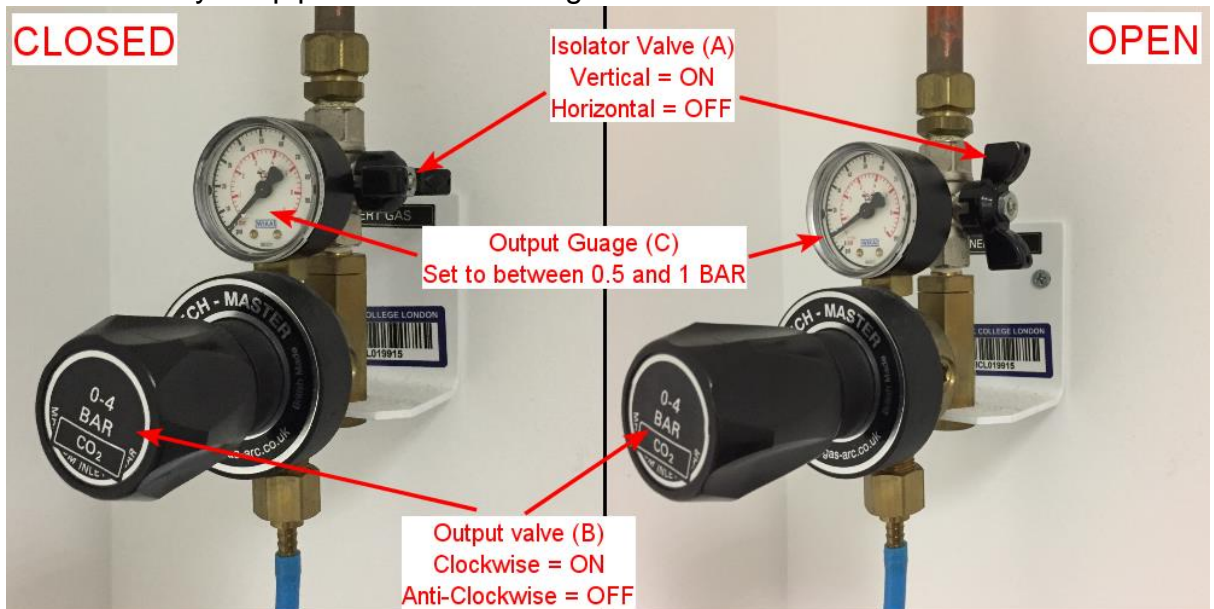


Turning on

- Check the level of distilled water in the humidifying aspirator bottle inside the incubator chamber - should be about 2 cm full and gas should be bubbling from the aspirator
- Insert sample carrier and place the lid you are using over your sample and connect to the aspirator bottle

Gas Safety Check

- Check gauge (C): (there may be some residual pressure from the last use)
 - output gauge should be between 0 and 1 BAR
- Check the main isolator valve (A) is closed – it should be in the horizontal position.
- Check the output regulator valve (B) is closed – almost fully anticlockwise (it should feel loose, but it will feel stuck if turned too far anticlockwise)
- Check visually the pipework from the regulator to CO₂ controller



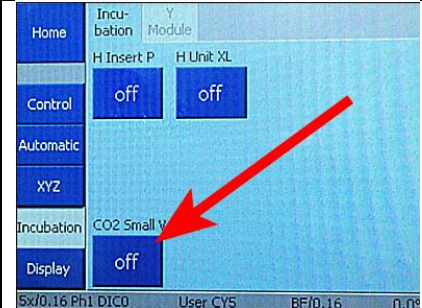
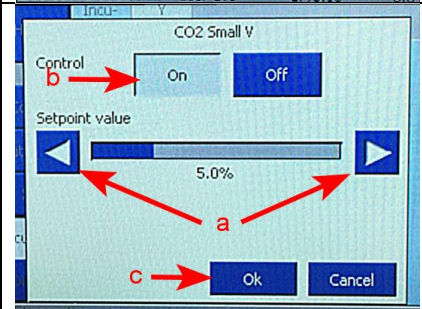
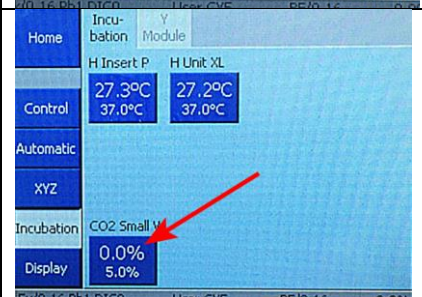
Turn on CO₂ gas

- Turn the main isolator valve (A) to the vertical position.
- Then by turning the output regulator valve (B) slowly clockwise, increase the pressure to between 0.5 and 1 BAR as displayed in the left hand (output) gauge - a slight resistance will be felt when the valve starts to open.
- **Important – do not set pressure above 1 bar or the tubing may separate.**

Microscope CO₂ Controller

Home	Objec- tives	Colibri LEDs	Re- flector	Opto- var	Light path
	5x	10x	20x	40x	Load position
Control	63x Oil	100x Oil			TL Illumination
Automatic					Off On
XYZ					RL Illumination
Incubation					Off On
Display					
5x/0.16 Ph1 DIC0 EGFP EGFP BF/0.09 0.0%					

- Start up the microscope in the usual way
- On the TFT screen, select “Microscope” and then “Incubation” (A) (if Incubation is not visible then re-start the system as above).

	<ul style="list-style-type: none"> • The display will show the CO2 Small V as OFF • To Switch on, press the OFF button to open the CO2 control panel
	<p>To adjust the CO2:</p> <ul style="list-style-type: none"> • Select the required temperature (a) and then press ON (b). Then press the OK button (c).
	<ul style="list-style-type: none"> • The display will now show the current and the required set CO2 values. • Press the Control button to return to the objective display

Check flow rate

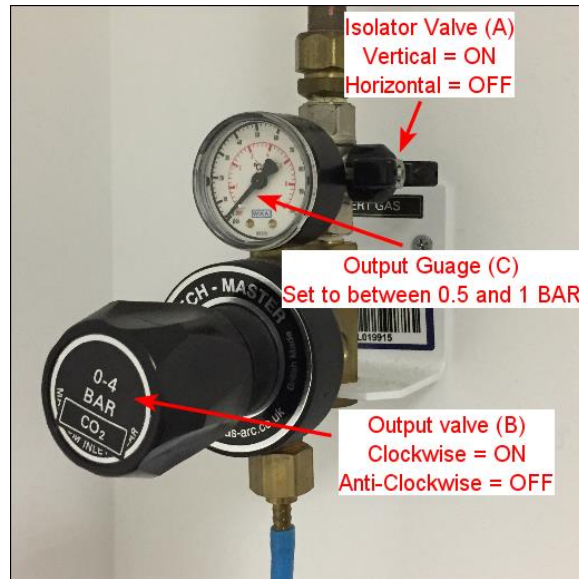
- Check for bubbling inside the aspirator bottles inside the incubator
- Check the output gauge again and adjust the output valve as necessary to maintain 0.5 to 1 BAR

Shutting down

CO₂

- Turn the main isolator valve (A) to the Horizontal closed position.

- Close the output regulator valve (B) – almost fully anticlockwise - it should feel loose, but it will feel stuck if turned too far anticlockwise



- Turn off CO₂ on the TFT controller to the right of the microscope

Heating

- Switch off the heating unit on the TFT controller
- Remove humidifier box if used